

# Hydrogeological Tracer Techniques

for Groundwater,  
Surface Water, and  
Mine Water

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South African Research Chair for Acid Mine Drainage Treatment

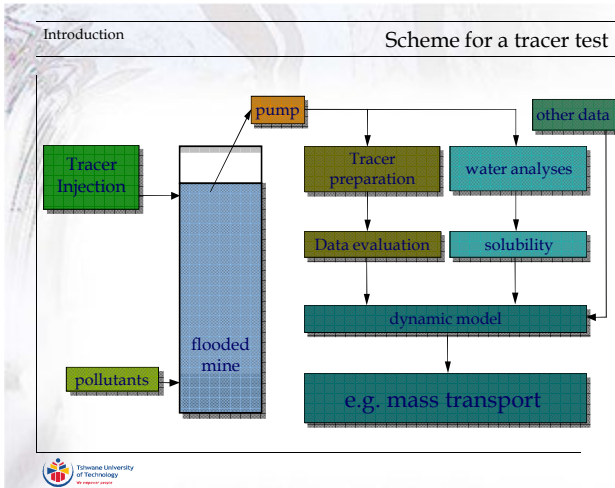
- Introduction
- Tracer types
  - Natural
  - Artificial
- Preparation of Tracer tests
- Legal Questions/Permission
- Implementation
- Evaluation
- Examples
  - Surface/Ground Water
  - Mine Water

- Hydraulic connections (flow paths)
- Ground water flow direction
- Ground water velocity, retention time
- Hydrodynamic dispersion
- Flow rate, mass transport
- Variability of hydrogeological properties
- Transport properties of dissolved and particulate water contents and their interactions

- Pollution of ground water
- Location and reliability of dams
- Mine water rebound after mine closure
- Flow of water within the flooded mine
- Drainage of contaminated mine water
- Duration of water treatment
- Pollution control by mine geometry
- Mass transport

- Transport with velocity of transport medium
- No natural occurrence of the tracer used
- Good analytical detection (even in big dilution)
- Water soluble or used in suspension
- High resistance (chemical stability)
- No interaction between tracer and medium
- Economic (reasonably priced according to buying, handling and analytical procedures)
- Physiological safe (non toxic)
- No lasting negative effects on (ground) water quality

- As deep as possible
- Injection without contamination
- More than one injection point
- Suitable tracer for mine water
- Tracer test must be repeatable
- Cheap and easy to handle



- Tracer Types Tracer types
- “Natural” Tracers
    - Environmental Isotopes
    - Environmental Chemicals
    - Organisms
    - Physical Effects
  - Artificial Tracers
    - Water Soluble
    - Water Insoluble
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- Tracer Types Natural Tracers
- Environmental Isotopes
    - Nuclides of substances, occurring in water
    - Usually without anthropogenic influence
    - Low concentrations
      - Expensive measuring techniques
      - Complicated interpretation
    - Huge spatial and temporal interpretation possible
    - Age or temperature determination
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Tracer Types Natural Tracers

- Environmental Isotopes (1/2)

Isotope	compound	half-life (years)	ratio
<sup>2</sup> H	<sup>2</sup> H <sup>1</sup> HO	stable	(9 – 17) · 10 <sup>-5</sup>
<sup>18</sup> O	H <sub>2</sub> <sup>18</sup> O	stable	(188 – 202) · 10 <sup>-5</sup>
<sup>3</sup> H	<sup>3</sup> H <sup>1</sup> HO	12.34	10 <sup>-16</sup> – 10 <sup>-17</sup>
<sup>3</sup> He	dissolved	stable	1.4 · 10 <sup>-6</sup>
<sup>4</sup> He	dissolved	stable	–
<sup>39</sup> Ar	dissolved	269	10 <sup>-15</sup>
<sup>85</sup> Kr	dissolved	10.76	6 · 10 <sup>-12</sup>

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Tracer Types Natural Tracers

- Environmental Isotopes (2/2)

Isotope	compound	half-life (years)	Ratio
<sup>36</sup> Cl	<sup>36</sup> Cl <sup>-</sup>	3.01 · 10 <sup>5</sup>	2 · 10 <sup>-12</sup>
<sup>13</sup> C	<sup>13</sup> CO <sub>2</sub> , H <sup>13</sup> CO <sub>3</sub> <sup>-</sup>	Stable	1.1 · 10 <sup>-2</sup>
<sup>14</sup> C	<sup>14</sup> CO <sub>2</sub> , H <sup>14</sup> CO <sub>3</sub> <sup>-</sup>	5,730	1.2 · 10 <sup>-12</sup>
<sup>34</sup> S	<sup>34</sup> SO <sub>4</sub> <sup>2-</sup>	Stable	4.2 · 10 <sup>-2</sup>
<sup>15</sup> N	<sup>15</sup> NO <sub>3</sub> <sup>-</sup>	Stable	3.7 · 10 <sup>-3</sup>
<sup>234</sup> U	<sup>234</sup> U <sup>6+</sup>	2.47 · 10 <sup>5</sup>	5 · 10 <sup>-3</sup>

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Tracer Types Environmental Isotopes

- Radioactive Decay (1/2)

$$n_t = n_0 e^{-\lambda t}$$

$n_t$ : number of atoms after time  $t$   
 $n_0$ : number of atoms at  $t = 0$   
 $\lambda$ : decay constant  
 $t$ : time

$$\lambda = \ln 2 (T_{1/2})^{-1} = 0.6931 \cdot (T_{1/2})^{-1}$$

$T_{1/2}$ : half-life time

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- Radioactive Decay (2/2)

$$\tau_a = T_{1/2} (\ln 2)^{-1} \ln (n_0 \cdot n_t^{-1})$$

$\tau_a$ : time during radioactive decay

$T_{1/2}$ : half-life time

$n_0$ : number of atoms at  $t = 0$

$n_t$ : number of atoms after time  $t$



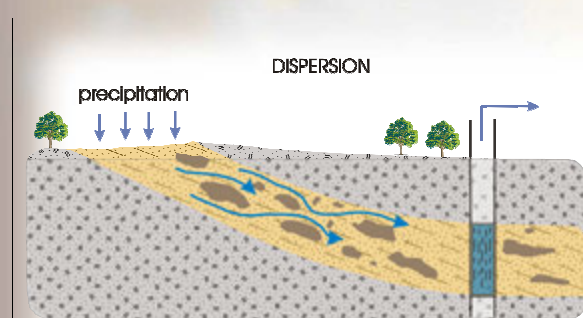
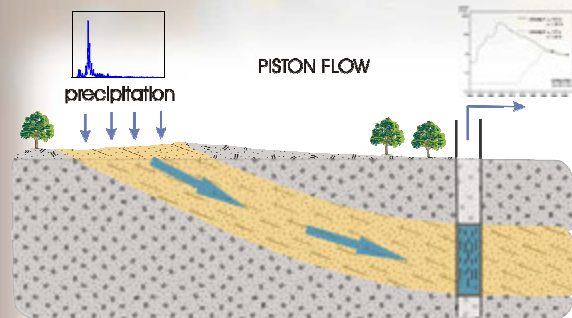
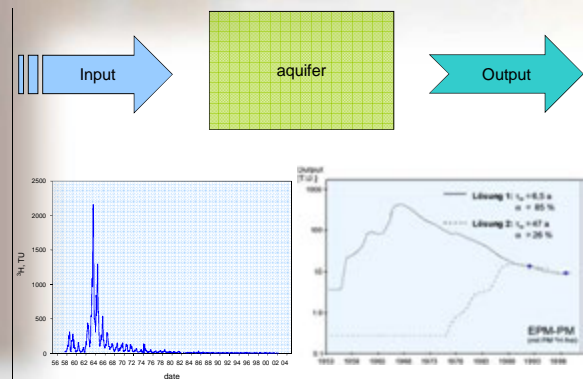
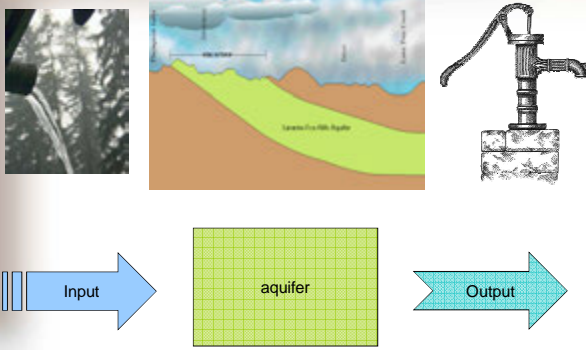
- Input - Output function

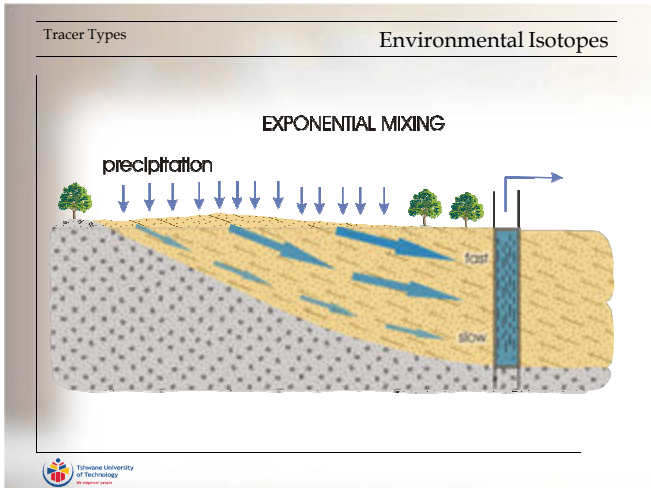
$$c_{out, t} = f(c_{in, t})$$

- PFM: Piston Flow Model

- EM: Exponential Model

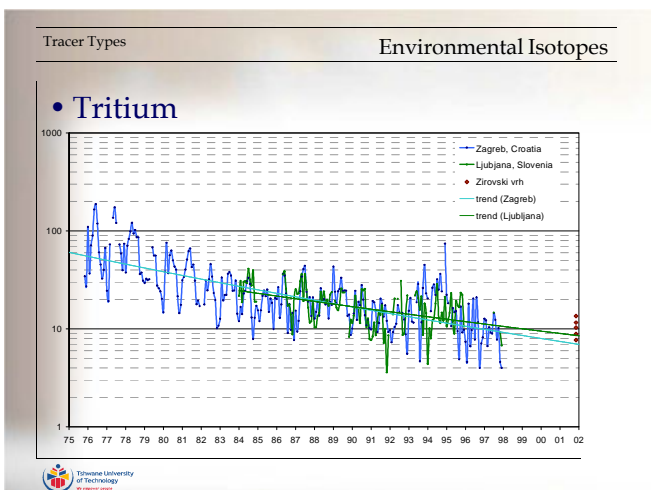
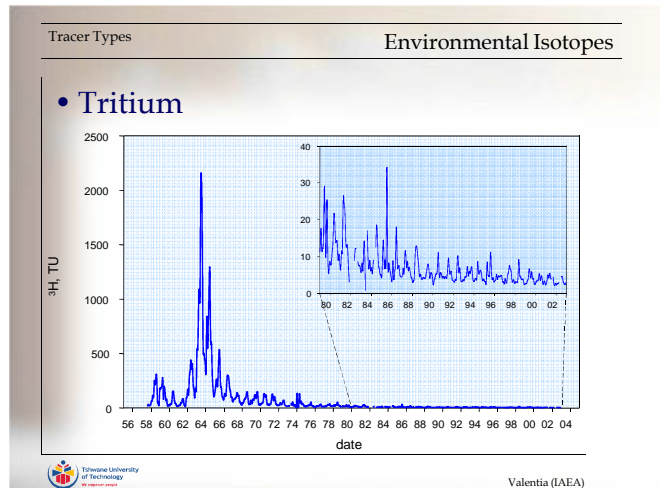
- DM: Dispersion Model



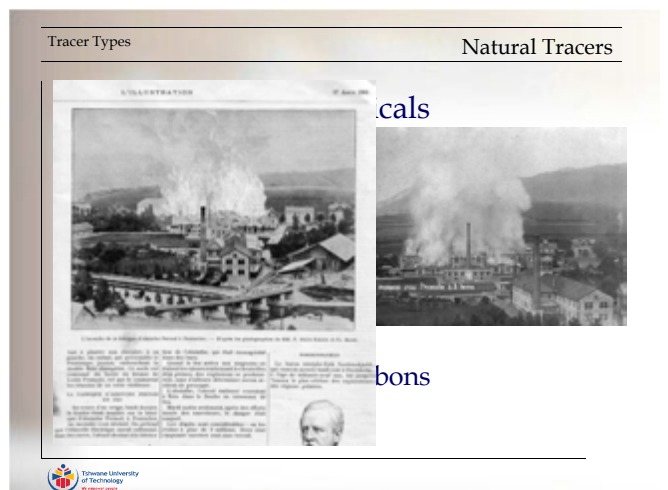
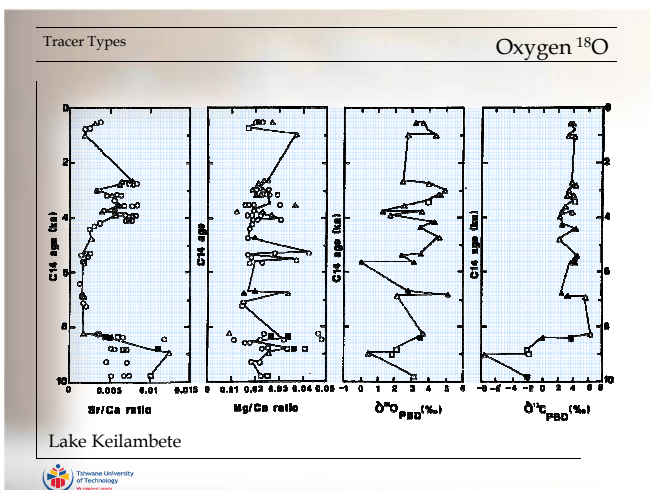
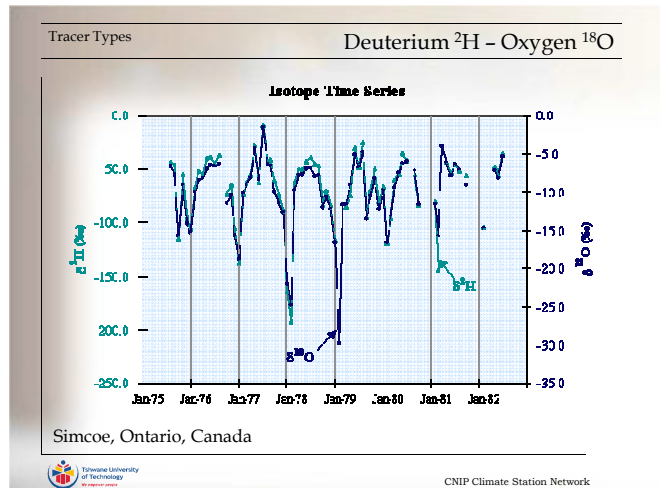
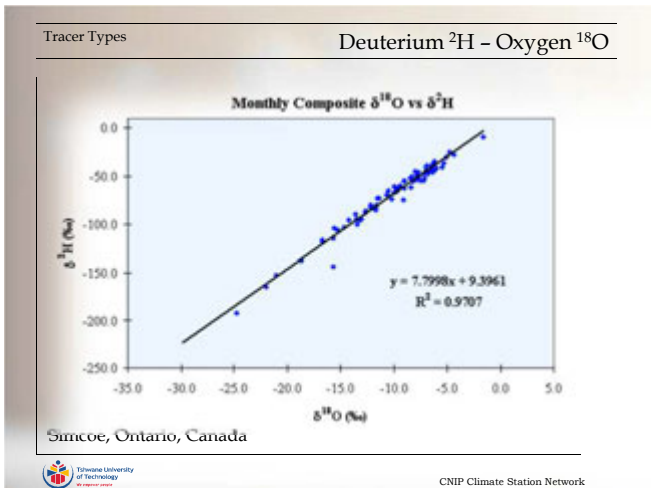
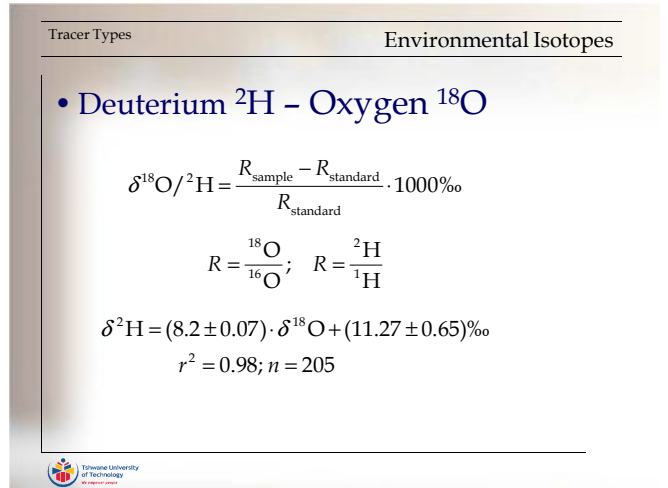
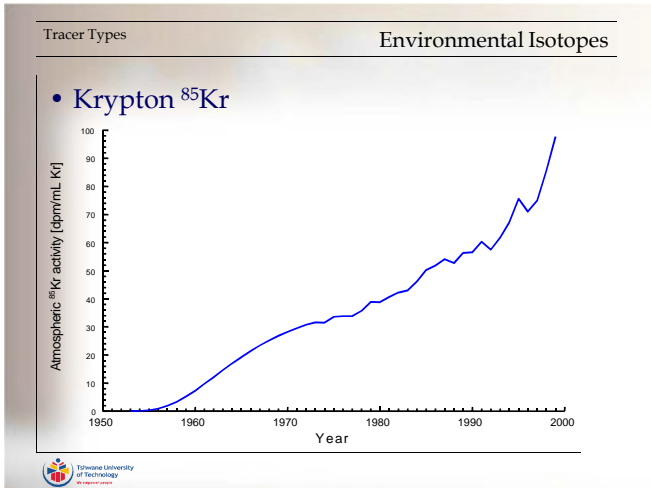


- Tracer Types Environmental Isotopes
- Tritium ( $^3\text{H}$ )
    - Natural  $^3\text{H}$ -content: < 10 TU
      - Stratospheric high energy protons ( $E > 100$  MeV) react with nitrogen and oxygen
    - $1 \text{ TU} \triangleq 0.118 \text{ Bq kg}^{-1}$
    - Nuclear weapons' tests (1952 – 1963)
    - Input function with peak between 1963 and 1965 (up to 10,000 TU)
    - seasonal variations (winter: low; summer: high)
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- Tracer Types Environmental Isotopes
- Tritium
    - several models for result's interpretation
      - PFM: Piston Flow Model
      - EM: Exponential Model
      - DM: Dispersion Model
    - today: between 10 and 30 TU
    - more  $^3\text{H}$  in northern than in southern hemisphere (mixing effect)
    - IAEA precipitation collection network
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- Tracer Types Environmental Isotopes
- Krypton  $^{85}\text{Kr}$ 
    - Emitted from nuclear plants
    - Quasi-linear input function since 1958
      - 1950-1958: 0 dpm/ml  $^{85}\text{Kr}$
      - 1980: 40 dpm/ml  $^{85}\text{Kr}$
      - 2000: 80 dpm/ml  $^{85}\text{Kr}$
    - Quick mixing in atmosphere
    - Low seasonal and local variations
    - water marked by  $^{85}\text{Kr}$  dissolved in air
    - No disturbing  $^{85}\text{Kr}$  in the underground
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- Uranine (2.5 – 2,400 mg L<sup>-1</sup>)

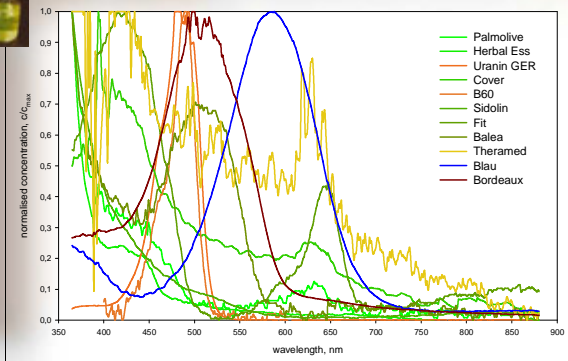
- Bath Products
- Cosmetics
- Cleaning Products



Badedas	2,400 mg L <sup>-1</sup>
Benny	410 mg L <sup>-1</sup>
Dressin Pine needle	1,350 mg L <sup>-1</sup>
General	15.5 mg L <sup>-1</sup>
Wundi	2.5 mg L <sup>-1</sup>



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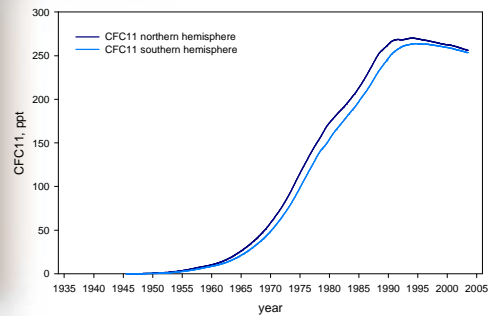


- Borate

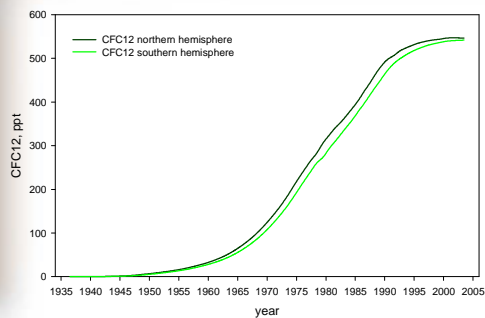
- Sewage
- Waste Deposits
- Laundry Detergents
- up to 83 mg L<sup>-1</sup> HBO<sub>2</sub>



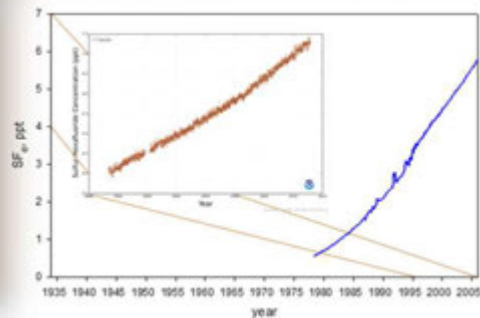
- Chlorinated Hydrocarbons / SF<sub>6</sub>



- Chlorinated Hydrocarbons / SF<sub>6</sub>



- Chlorinated Hydrocarbons / SF<sub>6</sub>



Tracer Types Natural Tracers

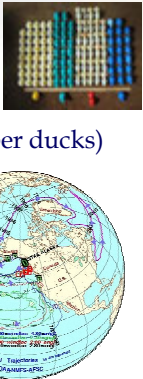
- Organisms
  - Bacteria
  - Pollen
  - Eels
  - Ducks



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Tracer Types Rubber Duckies

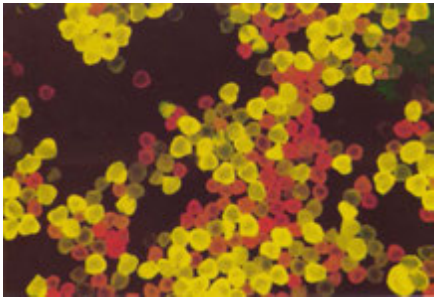
- Spill over: 10 January 1992
- 44.7°N, 178.1°E
  - North American west coast
- 29,000 bathtub toys (including rubber ducks)
- Ten months later the toys began washing ashore near Sitka, Alaska
- Ocean Surface Current Simulations (OSCURS)



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Tracer Types Organisms

- Pollen (*Lycopodium clavatum*)



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Tracer Types Natural Tracers

- Physical Effects
  - Temperature
    - Altitude effect
  - Salinity
    - Waste deposits
    - Salt lakes
    - Ocean influence

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Tracer Types Artificial Tracers

- Artificial Tracers
  - Water soluble
    - e.g. dyes
    - salts
  - Water insoluble
    - e.g. spores
    - microspheres

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Tracer Types Artificial Tracers: Water Soluble

- Dyes
  - Fluorescent dyes
  - Non-fluorescent dyes
- Salts
- Radioactive Tracers
- Activation analytical tracers

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- filtering of white light
  - dyes are coloured
- absorption can be measured
  - dye intensity
- fluorescent dyes
  - light energy is emitted
    - > Fluorescence
  - detection limit  $10^4$  times lower than for non-fluorescent dyes



- Characteristic wave lengths for each dye
  - Absorption spectra
  - Stimulation wave length
  - Emission spectra
- Important: wave length maxima
- Detection limit for dyes with two maxima is lower than with one maximum



- Chemical composition and structure
  - Organic molecules
    - C, H, O, N, S, Na, Cl, Br
  - Chemical structure, Arrangement of atoms results in colour and fluorescence
  - chemical structures must not be destroyed during the tracer test
  - fluorescence is a result of double bindings in the aromatic molecules
  - Xanthene-structure of most fluorescent dyes



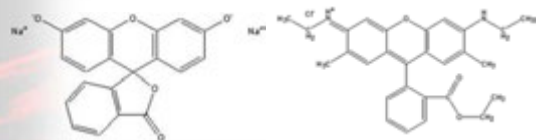
- Chemical composition and structure
  - Trivial names instead of exact chemical name
    - Fluorescein: 9-(2-carboxyphenyl)-3-hydroxy-6-fluoron
  - Be exact when using trivial names!
  - Use full trivial name (e.g. there are many rhodamins)
  - Use CAS number
    - Na-Fluorescein: 518-47-8
  - Fluorescein  $\neq$  Uranine



- Chemical composition and structure
  - Many names for one substance might be in use e.g.
    - uranine
    - c.i. acid yellow 73
    - fluorescein sodium; Na-fluorescein
    - spiro(isobenzofuran-1(3h),9'-(9h) xanthen)-3-one,3',6'-dihydroxy-, disodium salt (9ci)
    - Fluorescein (often used in Anglo-Saxon)
    - disodium salt
    - yellow 73, c.i. acid



- Chemical composition and structure



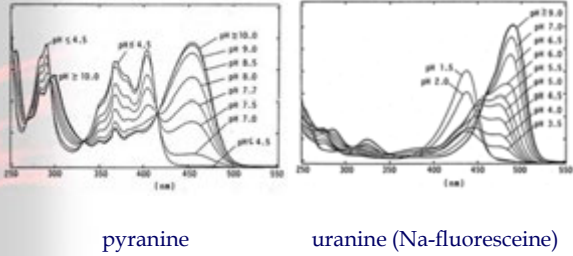
Uranine (Na-fluoresceine)

Rhodamine 6G





- pH-dependency of fluorescence



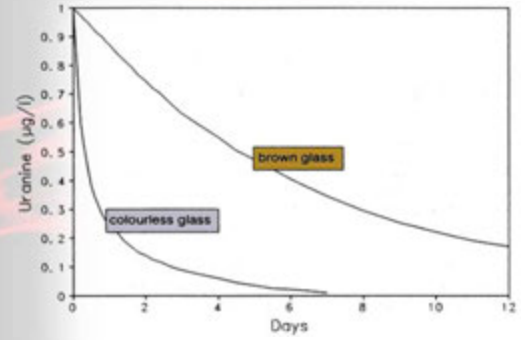
pyranine

uranine (Na-fluoresceine)



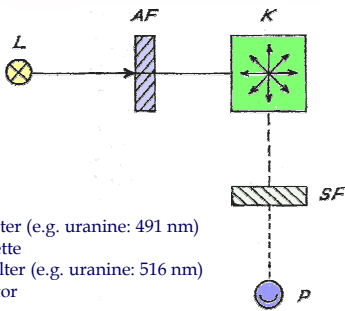
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- Decomposition of fluorescent tracers



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- Detection of fluorescent tracers

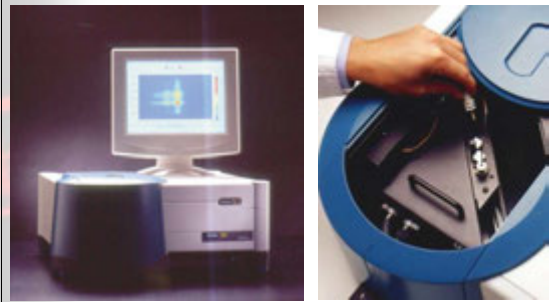


- L lamp
- AF excitation filter (e.g. uranine: 491 nm)
- K sample cuvette
- SF secondary filter (e.g. uranine: 516 nm)
- P photo detector



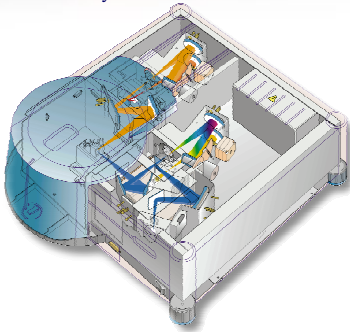
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- Modern Laboratory Fluorometer



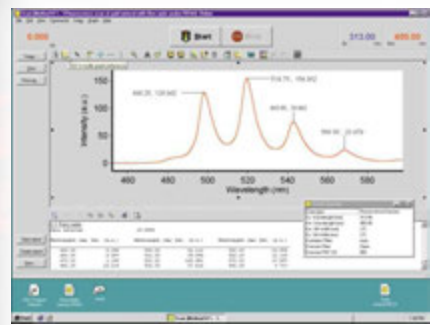
Varian Cary Eclipse

- Modern Laboratory Fluorometer



Varian Cary Eclipse

- Modern Laboratory Fluorometer



Varian Cary Eclipse

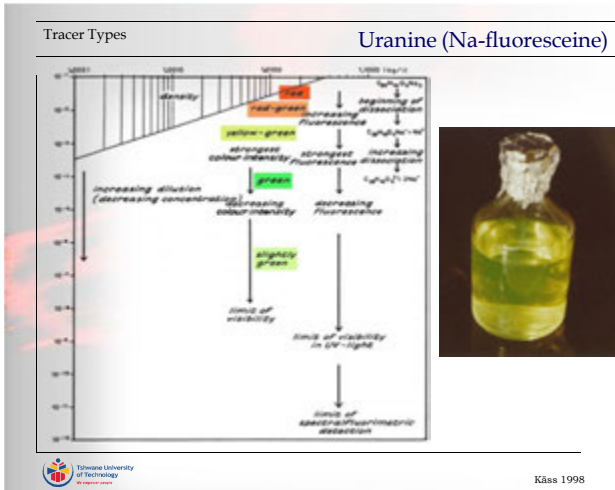
Tracer Types		Fluorescent Dyes	
Tracer	CAS number	Detection limit, µg/L	Sorptivity
Uranine, Na-fluorescein	518-47-8	0.002	very low
Eosin Y	17372-87-1	0.01	low
Pyranin	6358-69-6	0.008	very low
Sulforhodamine G	5873-16-5	0.005	weak
Sulforhodamine B	3520-42-1	0.007	weak
Rhodamine B	81-88-9	0.006	strong
Rhodamine WT	37299-86-8	0.006	medium
Lissamine FF	2391-30-2	0.03	medium
Tinopal CBS-X	38775-22-3	0.07	medium
Naphthionate	130-13-2	0.01	low



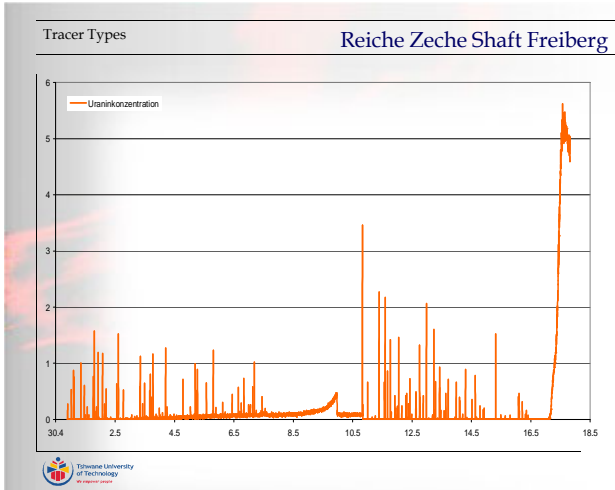
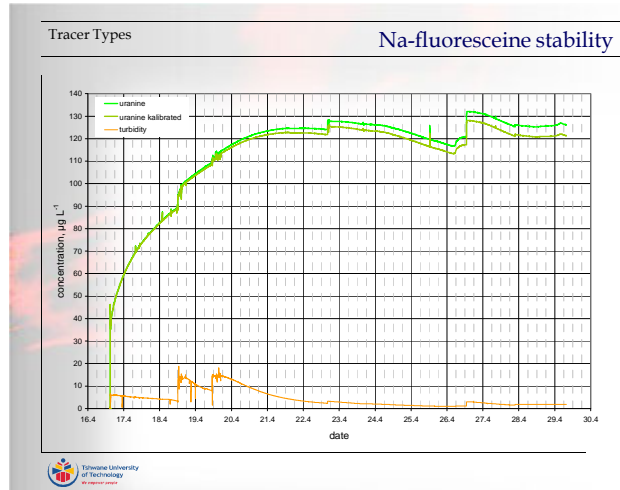
Tracer Types		Fluorescent Dyes	
Tracer	CAS-Number	Toxicological assessment	Assessment basis
Uranine (sodium fluorescein)	518-47-8	Safe	Tox, Lit
Eosin yellow (Eosin Y)	17372-87-1	Safe	Lit, Work
Sulforhodamine B	3520-42-1	Ecotoxicologically unsafe	Tox
Amidiorhodamine G	5873-16-5	Safe	Tox
Rhodamine WT	37299-86-8	Not recommended	Tox
Rhodamine B	81-88-9	Not recommended	Tox, Lit
Rhodamine 6G	989-38-8	Not recommended	Tox, Lit
Sodium naphthionate	130-13-2	Safe	Tox
Pyranine	6358-69-6	Safe	Tox
Tinopal CBS-X	38775-22-3	Safe	Tox
Tinopal ABP liquid	68155-70-4	Safe	Tox
Lithium salts		Safe with restrictions	Lit, Work
Strontium salts		Safe with restrictions	Lit, Work
Bromides		Safe with restrictions	Lit, Work
Activatable isotopes		Safe with restrictions	Lit, Work
microspheres	9003-53-6	Safe	Lit, Work
Hub moss spores	-	Safe	Tox, Work



Modified from Behrens et al. 2001, CAS numbers added



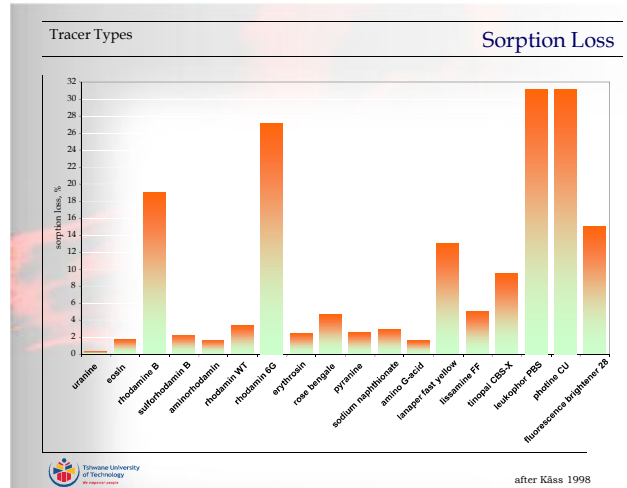
Käss 1998



Georgi Unterbau, Bridlegg

Tracer Types Na-fluoresceine

Stöckelgraben Speyer 15.11.2002



Tracer Types Non-fluorescent Dyes

☹️

- poor detection limits
- only small surface of water body
- short travel distances
- water of good quality

☺️

- good light and pH stability
- modern spectral photo-meters can be used

Tracer Types Non-fluorescent Dyes

- Indigo (Riba Reka, Slovenia 1864)
- Aniline red (Danube 1869)
- Fuchsine (Lur cave system 1928)
- Congo red, malachite green (Mendip Hills 1968)
- Brilliant blue (1994, 1995)
- Aniline violet
- Auramine
- Methyl blue

Tracer Types Salts

- NaCl
- LiCl
- KCl
- Borax ( $\text{Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O}$ )
- SrCl
- CsCl
- Br<sup>-</sup>
- I<sup>-</sup>
- Ammonium
- Dichromate
- ZnCl<sub>2</sub>

Tracer Types Salts: Introduction

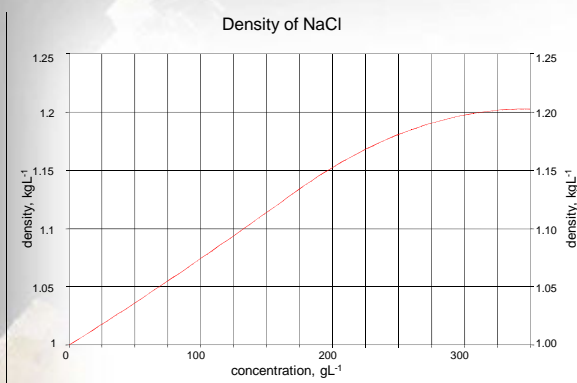
- first test 1872 (900 kg near Lausen, Switzerland)
- first important use of salts in 1877
  - Danube Immendingen
  - NaCl: 10,000 kg (+ 10 kg Na-Fluorescein)
  - recovery 92.6 %
- in natural or mine waters often high background of "Salts"
  - Gernrode/Harz: 1,700 mgL<sup>-1</sup> Cl<sup>-</sup>
  - Niederschlema/ Alberoda: 5.6 mgL<sup>-1</sup> Li<sup>+</sup>
- always measure background concentrations
- also: sewage water; landfill drainage water; polluted rivers

- high salt concentrations might be harmful for biology and microbiology
  - trees and/or worms might die
- increase in electrical conductivity can be used to determine salt concentration
  - used in surface flow investigations
  - high amounts of salt necessary
  - in practice for NaCl only
- salts: cation + anion
- usually, only one ion is tracer
- high density of salts can result in density flows

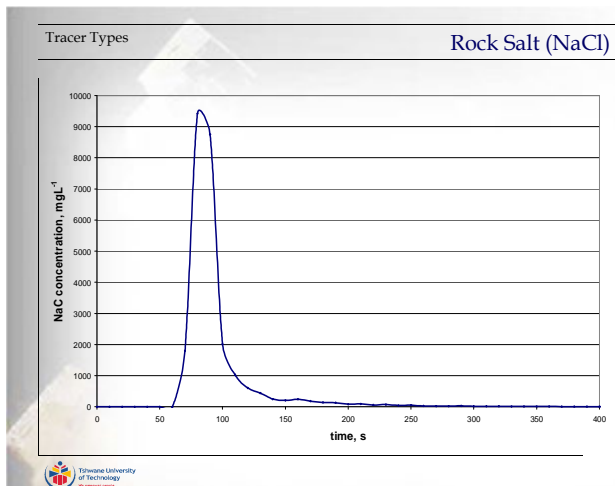
- cations and anions show different flow characteristics
  - example: LiBr
  - either use LiCl, KBr, or NaBr
- adsorption or cation exchange might cause loss of ions
  - higher in pore than in karst water
  - ion exchange capacity falls with increasing ion potential
  - Li: ionic potential 1.28  $\Rightarrow$  low exchange capacity
  - Cs: ionic potential 0.61  $\Rightarrow$  high exchange capacity

- electrical conductivity
  - mainly NaCl
- titration of Cl<sup>-</sup> anion
- flame emission photometry
- atom absorption spectrophotometry
- photometry
  - e.g. permanganate
- HPLC + ion chromatography
  - Br, I
- ICP-MS
  - Inductively Coupled Plasma - Mass Spectrometry

- first and most widely used salt tracer
- dissociation into Na<sup>+</sup> and Cl<sup>-</sup>
- samples should always be analysed for both ions
- normally, the Cl<sup>-</sup> is the tracer
- saturation: 0 °C 356 gL<sup>-1</sup>; 30 °C 361.5 gL<sup>-1</sup>
- to avoid sinking of the solution:
  - warm water
  - max concentration 0.5 % NaCl (3 gL<sup>-1</sup>)
- sample volume at least 250 mL when using titration of Cl<sup>-</sup>



- detection methods
  - gravimetical (AgCl)
  - titrimetical (NaCr<sub>2</sub> + AgCl)
  - flame emission at 589.3 nm
  - ion sensitive electrode (not known for chloride determination on site)
  - electrical conductivity (high precision)
  - ion chromatography (only with low concentrations)
- toxicology (LD<sub>50</sub>):
  - warm blooded animals: 3,000 mg kg<sup>-1</sup>
  - fish and fish fry: 10,000 mgL<sup>-1</sup>



- Tracer Types Lithium-salts
- first use in 1907
    - 50 kg in Notranjska Reka near St. Kanzian, Slovenia
  - Li has a small ionic radius, therefore least subjected to ion exchange
  - possible use in porous aquifers with flow distances below ca. 200 m
  - cheapest Li-Salt is LiCl (approx. € 10  $\text{kg}^{-1}$ )
  - technical purity grade (99 %) is sufficient
  - solubility: 20 °C 832  $\text{g L}^{-1}$ ; 60 °C 984  $\text{g L}^{-1}$
  - exothermic reaction during solution  $\Rightarrow$  avoid plastic containers
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- Tracer Types Lithium-salts
- background Li and variation must be known
  - detection methods
    - Ca has to be removed prior to analyses to increase Li detection limits (ammonium oxalate)
    - flame photometry
    - ICP-MS
    - bad: atomic absorption spectrometry (AAS)
  - toxicology ( $\text{LD}_{50}$ ):
    - warm blooded animals: 526  $\text{mg kg}^{-1}$
    - fish and fish fry: 2,000 – 3,500  $\text{mgL}^{-1}$
    - no more than 0.5  $\text{mg L}^{-1}$  at public water supplies during a tracer test
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- Tracer Types KCl
- Only used when K should be the tracer
    - K concentrations are lower than Na concentrations (0.1 – 8  $\text{mgL}^{-1}$ )
    - Na and K have approx. same abundance in crust
    - K is bound in rock building minerals (e.g. feldspar)
  - If Cl is meant to be the tracer the cheaper NaCl should be used (cost KCl = 4  $\times$  cost NaCl)
  - K is more sorptive and subject to ionic exchange than Na
  - In tracer tests, exchanged Ca often arrives before K
- Tishane University of Technology

- Tracer Types KCl
- solubility
    - 10 °C: 313  $\text{gL}^{-1}$
  - detection methods
    - flame emission
    - ionic chromatography
    - not advisable: AAS (766.5 nm)
  - toxicology:
    - Humans (t): 2,430  $\text{mg kg}^{-1}$
    - fish (t): 700 – 5,200  $\text{mgL}^{-1}$
    - water organisms (t): 400 – 5,000  $\text{mgL}^{-1}$
- Tishane University of Technology

- Tracer Types Borax ( $\text{Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O}$ )
- first used in 1948/49 (Oilfields in Kansas and Oklahoma)
  - transported as undissociated form
    - $\Rightarrow$  low adsorption
    - $\Rightarrow$  ion exchange
    - Only clay minerals can adsorb small amounts of B
  - natural B concentration of water must be known (0.02 – 121  $\text{mgL}^{-1}$  observed)
  - don't warm water to dissolve Borax (recrystallisation)
  - adding up to 30 % of boric acid increases the dissolution
  - Unimportant in artificial tracer tests but as natural tracer (sewage water, waste deposits)
- Tishane University of Technology

Tracer Types	Borax ( $\text{Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O}$ )
<ul style="list-style-type: none"> <li>solubility           <ul style="list-style-type: none"> <li>– 10 °C: 16.2 gL<sup>-1</sup>; 20 °C: 25.2 gL<sup>-1</sup></li> </ul> </li> <li>detection methods           <ul style="list-style-type: none"> <li>– colorimetric               <ul style="list-style-type: none"> <li>• B + Azomethin H ⇒ yellow dye</li> </ul> </li> <li>– ICP-MS</li> <li>– fluorimetric boric acid determination</li> </ul> </li> <li>toxicology (LD<sub>50</sub>):           <ul style="list-style-type: none"> <li>– rats: 4,500–5,000 mg kg<sup>-1</sup></li> <li>– Humans (†): 500–5,000 mg kg<sup>-1</sup></li> <li>– fish (†): 3,000–7,000 mgL<sup>-1</sup></li> <li>– water organisms (†): 540–9,900 mgL<sup>-1</sup></li> </ul> </li> </ul>	

Tracer Types	SrCl <sub>4</sub> ·6H <sub>2</sub> O
<ul style="list-style-type: none"> <li>Sr, not Cl is the tracer</li> <li>natural Sr concentration (“background”) must be known (0.005–14 mgL<sup>-1</sup>)</li> <li>compared to Na-fluoresceine, a retardation factor of 10 must be expected</li> <li>Amounts used so far: 50 kg; 2 kg</li> <li>detection methods           <ul style="list-style-type: none"> <li>– analyse within 48 hours to avoid Sr precipitation!</li> <li>– Flame emission</li> <li>– AAS</li> <li>– ICP-MS</li> </ul> </li> <li>toxicology:           <ul style="list-style-type: none"> <li>– trout (†): 10,000 mgL<sup>-1</sup></li> </ul> </li> </ul>	

Tracer Types	CsCl
<ul style="list-style-type: none"> <li>used in Notranjska Reka (Slovenia) and Upper Rhine Valley</li> <li>highly sorptive!</li> </ul>	

Tracer Types	Bromide ( $\frac{1}{2}$ )
<ul style="list-style-type: none"> <li>only present in traces in natural waters</li> <li>no sorption</li> <li>stable towards microbiological activities: but uptake by plants</li> <li>but: high costs and complicated analytical procedures</li> <li>good water solubility of NaBr           <ul style="list-style-type: none"> <li>– 10 °C: 850 gL<sup>-1</sup></li> </ul> </li> <li>detection methods           <ul style="list-style-type: none"> <li>– ion sensitive electrodes               <ul style="list-style-type: none"> <li>• high detection limit: 50 µgL<sup>-1</sup></li> </ul> </li> <li>– ion chromatography</li> <li>– isotope dilution analyses (change of isotope ratios)</li> </ul> </li> </ul>	

Tracer Types	Bromide ( $\frac{2}{2}$ )
<ul style="list-style-type: none"> <li>not to be used near drinking water supplies, if water is chlorinated or ozonated           <ul style="list-style-type: none"> <li>– bromate and Br-organics will be formed</li> </ul> </li> <li>toxicology (LD<sub>50</sub>)           <ul style="list-style-type: none"> <li>– Rat: 3,500 mg/kg</li> <li>– Trout, bluegill, sunfish (96 h): &gt; 1000 mgL<sup>-1</sup></li> <li>– Water flea (48 h): &gt; 1000 mgL<sup>-1</sup></li> </ul> </li> </ul>	

Tracer Types	Iodide
<ul style="list-style-type: none"> <li>smaller natural concentrations than Br</li> <li>iodite can be chemically or microbiologically affected</li> <li>only suitable for           <ul style="list-style-type: none"> <li>– short flow distances</li> <li>– short residence times</li> <li>– low organic pollution</li> </ul> </li> <li>detection methods           <ul style="list-style-type: none"> <li>– ion sensitive electrodes</li> <li>– ion chromatography</li> <li>– isotope dilution analyses (change of isotope ratios)</li> </ul> </li> </ul>	

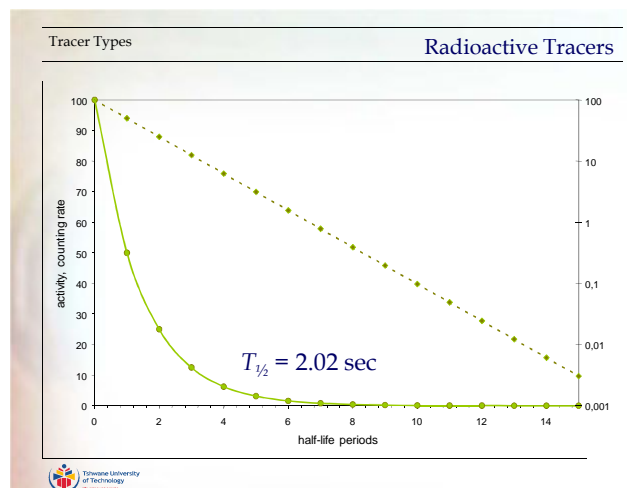
Tracer Types	Ammonium
<ul style="list-style-type: none"> <li>• first use 1942 (Warsteiner Massenkalk)</li> <li>• sensitive to sorption and oxidation</li> <li>• too much amounts must be used               <ul style="list-style-type: none"> <li>- e.g. 72 kg Warstein tracer test</li> <li>- <math>\Rightarrow</math> restricted use</li> </ul> </li> <li>• toxicology               <ul style="list-style-type: none"> <li>- in alkaline waters, ammonium is strongly toxic to fish</li> <li>- in strong sunshine pH can increase to 8, even 10</li> <li>- trout (†): 1.25–5 mgL<sup>-1</sup></li> </ul> </li> </ul>	

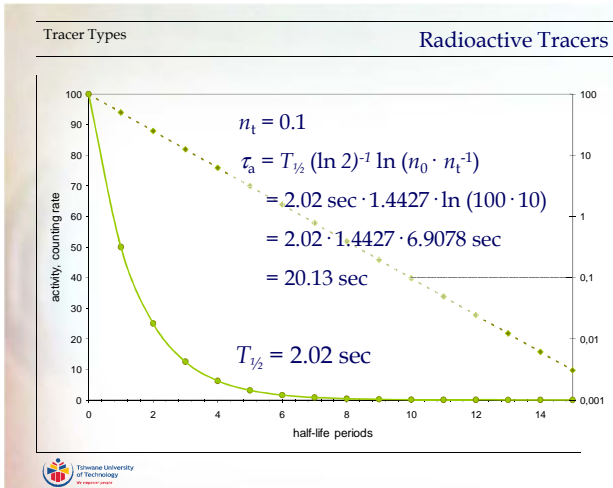
Tracer Types	Dichromate (Cr <sub>2</sub> O <sub>7</sub> <sup>2-</sup> )
<ul style="list-style-type: none"> <li>• discharge measurements in channels and rivers (up to 50 m<sup>3</sup> s<sup>-1</sup>)</li> <li>• 2 kg per m<sup>3</sup> s<sup>-1</sup> of flow needed</li> <li>• solubility               <ul style="list-style-type: none"> <li>- K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub>: 77 gL<sup>-1</sup> (10 °C)</li> <li>- Na<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub>·2H<sub>2</sub>O: 1,700 gL<sup>-1</sup> (10 °C)</li> </ul> </li> <li>• detection methods               <ul style="list-style-type: none"> <li>- photometric</li> <li>- colorimetric with diphenylcarbazide</li> </ul> </li> <li>• toxicology               <ul style="list-style-type: none"> <li>- very toxic!</li> <li>- trout (†): 0.2–0.35 mgL<sup>-1</sup></li> <li>- drinking water: 0.05 mgL<sup>-1</sup></li> </ul> </li> </ul>	

Tracer Types	Radioactive Tracers
• <sup>3</sup> H	12.3 a <sup>3</sup> H <sup>2</sup> HO     β
• <sup>24</sup> Na	15.0 h     Na <sup>+</sup> -cation     β
• <sup>51</sup> Cr	27.7 d     EDTA-complex     γ
• <sup>58</sup> Co	70.8 d     [Co(CN) <sub>6</sub> ] <sup>3-</sup> γ
• <sup>82</sup> Br	36 h     Br <sup>-</sup> -anion     γ
• <sup>85</sup> Kr	10.76 a     gas     γ
• <sup>114</sup> In	49.5 d     EDTA-complex     γ
• <sup>131</sup> I	8.05 d     I <sup>-</sup> -anion     γ
• <sup>198</sup> Au	2.7 d     Au <sup>3+</sup> -cation     β

Tracer Types	Radioactive Tracers
<ul style="list-style-type: none"> <li>• <b>Radioactive Decay (1/2)</b></li> </ul> $n_t = n_0 e^{-\lambda t}$ <p> <math>n_t</math>: number of atoms after time <math>t</math>  <math>n_0</math>: number of atoms at <math>t = 0</math>  <math>\lambda</math>: decay constant  <math>t</math>: time         </p> $\lambda = \ln 2 (T_{1/2})^{-1} = 0.6931 \cdot (T_{1/2})^{-1}$ <p><math>T_{1/2}</math>: half-life time</p>	

Tracer Types	Radioactive Tracers
<ul style="list-style-type: none"> <li>• <b>Radioactive Decay (2/2)</b></li> </ul> $\tau_a = T_{1/2} (\ln 2)^{-1} \ln (n_0 \cdot n_t^{-1})$ <p> <math>\tau_a</math>: time during radioactive decay  <math>T_{1/2}</math>: half-life time  <math>n_0</math>: number of atoms at <math>t = 0</math>  <math>n_t</math>: number of atoms after time <math>t</math> </p>	





- Tracer Types Activable Isotope Tracers
- substances must be under detection limit in the water to be tested
  - representative tracer properties
  - most important activation process: neutron activation
  - nuclides formed mostly unstable
  - sensitivity increases with increasing radioactivity
  - atomic reactors have to be used (e.g. Karlsruhe, Munich)
  - low detection limits
  - expensive method
- Tübingen University of Technology

- Tracer Types Activable Isotope Tracers
- $n, \gamma$ -process
    - $^{115}\text{In}$  (stable)  $\rightarrow$   $^{116}\text{In}$  ( $T_{1/2} = 54 \text{ min}$ )
  - $n, p$ -process
    - $^{32}\text{S}$  (stable)  $\rightarrow$   $^{32}\text{P}$  ( $T_{1/2} = 14.2 \text{ d}$ )
  - $n, \alpha$ -process
    - $^6\text{Li}$  (stable)  $\rightarrow$   $^3\text{H}$  ( $T_{1/2} = 12.3 \text{ a}$ )
  - detection limit increases with irradiation period
  - e.g. detection limit In: 0.0005 ppb
- Tübingen University of Technology

- Tracer Types Artificial Tracers: Water Insoluble
- drift particles
    - club moss spores (*Lycopodium clavatum*)
    - microparticles (microspheres)
    - bacteria
    - phages
    - geobombe
    - saw dust
    - tennis balls
    - yeast
    - eels
- Tübingen University of Technology

Tracer Types Club Moss Spores

- *Lycopodium clavatum*
- 50  $\mu\text{m}$  diameter
- first used in karst tracing
- natural and artificial colours possible

Tübingen University of Technology

Tracer Types Club Moss Spores

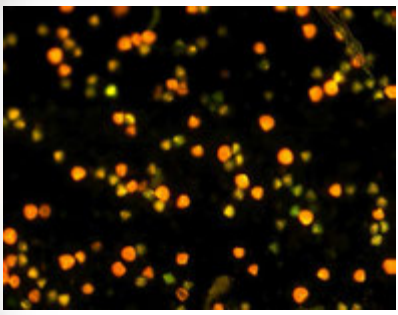
- Canadian *Lycopodium clavatum*

Tübingen University of Technology



Tracer Types Club Moss Spores

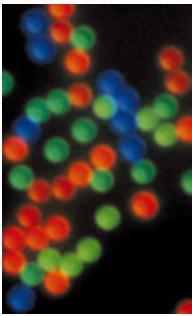
- Canadian *Lycopodium clavatum*



Tishreen University of Technology

Tracer Types Microparticles (Microspheres)

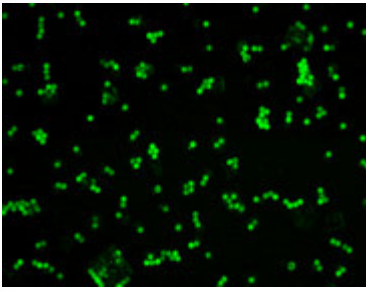
- microparticles (microspheres)
- 0.2 – 100  $\mu\text{m}$  diameter
- density 1.006  $\text{g cm}^{-3}$
- polystyrene
- latex
- positively, negatively, neutrally charged surfaces



Tishreen University of Technology

Tracer Types Microparticles (Microspheres)


- microparticles (microspheres)



Tishreen University of Technology

Tracer Types Microparticles (Microspheres)

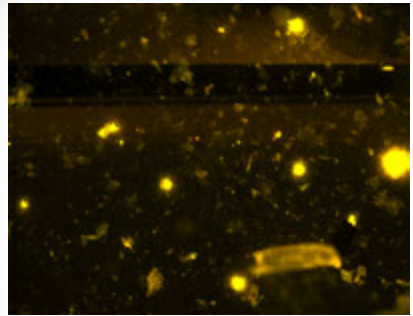
- microparticles (microspheres)



Tishreen University of Technology

Tracer Types Microparticles (Microspheres)

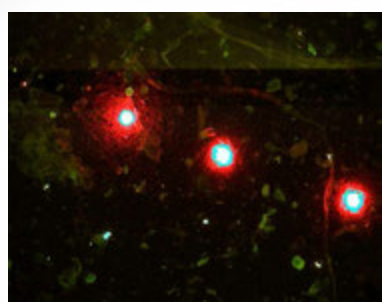
- microparticles (microspheres)



Tishreen University of Technology

Tracer Types Microparticles (Microspheres)

- microparticles (microspheres)



Tishreen University of Technology

Tracer Types Microparticles (Microspheres)

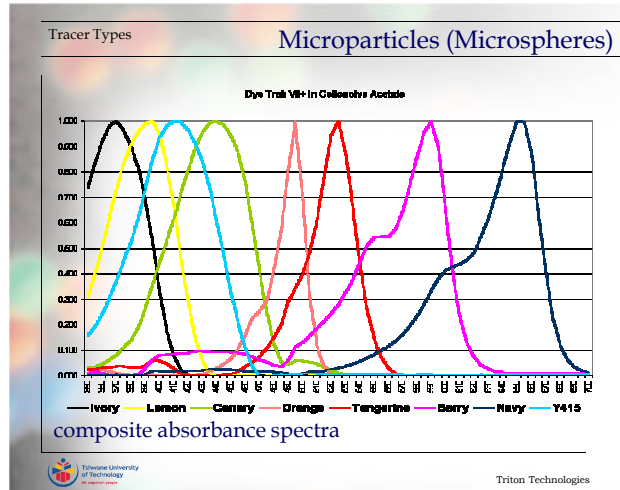
- microparticles (microspheres)



Blue      Yellow-Green      Red      Far Dark Red


Blue-Green      Orange      Crimson

Triton Technologies



Tracer Types Bacteria

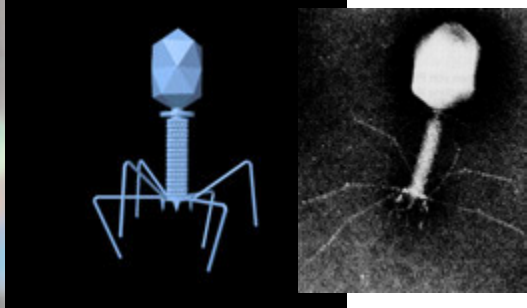
- bacteria (*Escherichia coli*)



Woods Hole Biological Laboratory

Tracer Types Phages

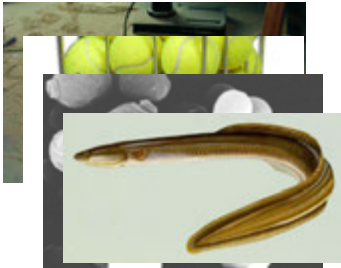
- phages (T4)



Triton Technologies

Tracer Types "Rarities"

- geobombe
- sawdust
- tennis balls
- yeast
- eels



Triton Technologies

Preparation of Tracer Tests Preparation of Tracer tests

- What data is necessary?
- Where shall the tracer be injected and sampled?
- Which tracer to be used?
- What amount to be injected?
- Who is responsible for what?
- ⇓
- Plan of the Tracer Test
- Documentation of Tracer Test

Triton Technologies

- always start with a
  - geological (e.g. rock types; sediments)
  - hydrogeological (e.g. type of aquifer, isolines: ground water contour lines)
  - hydrological (e.g. discharge; precipitation)
  - geochemical (e.g. anomalies)
  - hydrogeochemical (e.g. background values)
  - biological (e.g. bacteria, micro-organism)
- Investigation



- hydrogeological investigation
  - (ground) water flow direction
  - Isolines (ground water contour maps)
  - (ground) water velocity
  - dissolution of tracer
  - water usage
  - physico-chemical parameters
  - hydrogeochemistry



- Preparation of tracer tests needs time
- Go Out!
- Don't sit at your desk!



- Costs (direct)
  - Tracer
  - Sampling containers
  - Preliminary investigations
  - Preliminary expert reports
  - Site visits
  - Man power
  - Tracer analyses
  - Final report



- Costs (indirect)
  - Authorization (permits)
  - Sampling
  - Storage, transport of samples
  - Preparation of observation sites
  - Possible damages



- one or more injection sites
- based on preliminary investigations
- keep in mind:
  - you have to carry the injection equipment and tracers
  - situation could change between site visit and final test
- injections sites should be hydraulically connected to sampling sites
  - exception: unsaturated zone





- one or more sampling/observation sites
- better too much than too few of them
- you must be able to reach your sampling points in due time
- nearby sites could be hydraulically separated from each other
- worst thing that could happen:
  - negative result due to incorrect selection of injection and sampling sites!
- be open to any surprise!

- some tracers are light sensitive
- avoid sampling in polluted water
- don't sample treated water
- if necessary drill a borehole
- water with suspended matter should be filtered



- type of aquifer and lithology
- toxicology
- sorption and ion exchange
- pore sizes of aquifer
- water quality
- light impact
- water usage (e.g. fishing water)
- analytical criteria
- cost aspects
- injection site
- sampling site

- most tracer test guides: experience necessary
- EHTD 2003 (Efficient Hydrologic Tracer-Test Design): calculation is possible
- several equations used (EHTD lists 33 equations):
  - Dienert (1913): Na-fluoresceine (different aquifers)
  - Martel (1921): Na-fluoresceine (karst)
  - Timeus (1926): Na-fluoresceine (karst)
  - Aley & Fletcher (1976): Na-fluoresceine (karst, fissured)
  - UNESCO 1, 2 (1973/1983): Na-fluor. (porous/fissured)
  - Leibundgut & Wernli (1982): Na-fluoresceine (karst)
  - Käss (1992): general equation

- Käss (1992):  
 $M = L \cdot k \cdot B$   
 $L = 2.4 \text{ km}; k = 1; B = 3 \rightarrow$  Na-fluoresceine; porous aquifer  
 $M \approx 7 \text{ kg Na-fluoresceine}$
- UNESCO 1:  
 $M = L \cdot K$   
 $L = 2.4 \text{ km}; K = 1.5 \rightarrow$  Na-fluoresceine; porous aquifer  
 $M \approx 3 \text{ kg Na-fluoresceine}$



- e.g. Na-fluoresceine; porous aquifer  
 $Q = 20 \text{ L s}^{-1}; v = 10 \text{ m d}^{-1}; t = 3600 \text{ h}; L = 0.5 \text{ km}; s_1 = 100 \text{ m};$   
 $V = 100,000 \text{ m}^3; h = 10 \text{ m}; C_{\text{max}} = 10 \mu\text{g L}^{-1}; A = A_m = 1; K = 0.25 - 1$

equation	M, kg	variables
UNESCO 1	0.25–1	$K, L$
UNESCO 2	3	$K, Q, L, v, V$
BENDEL 1949	2.5	$K, Q, L$
LEIBUNDGUT 1974	1.3	$Q, t, C_{\text{max}}, A, „S“$
LEIBUNDGUT & WERNLI 1974	2.92	$t, h, s, C_{\text{max}}, A_m$



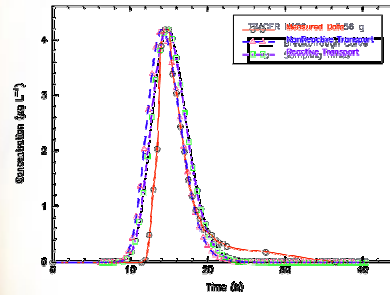
- e.g. WOLKERSDORFER et al. (2002):  
 spores, closed space (flooded mine)

$$m = \frac{1}{55,000} \times \frac{V_m}{r_r \times q} \times f$$

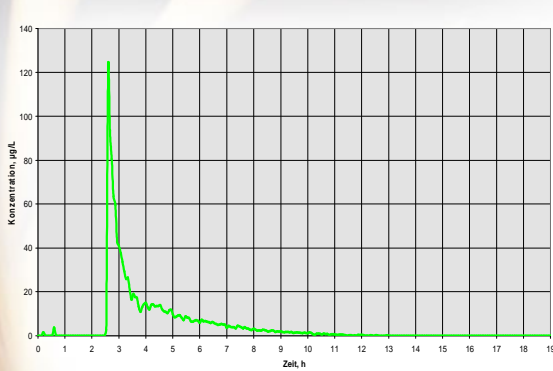
$m$ : mass/volume of tracer, g/mL;  $V_m$ : volume of enclosed fluid,  $\text{m}^3$ ;  $r_r$ : expected recovery rate (0.02–1);  
 $q$ : pumping capacity,  $\text{L min}^{-1}$ ;  $f$ : factor (spores 1; microspheres 0.25),  $\text{g L m}^{-3} \text{ min}^{-1}$



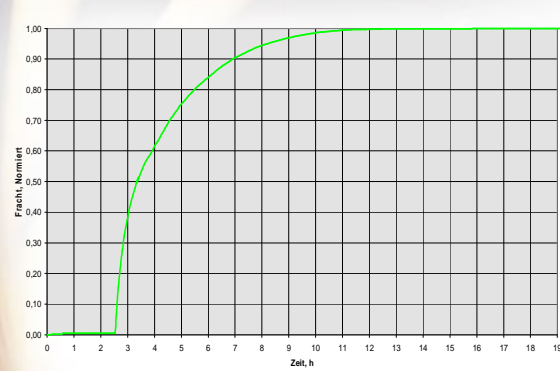
- EHTD: Efficient Hydrologic Tracer-Test Design  
 flowing stream; solution conduit (similar to “mine aquifer”)



Tracertest Reiche Zeche



Tracertest Reiche Zeche



- company
  - contract, money, site
- authorities
  - permits, public information
- you
  - everything – especially a good mood
- your employees
  - injection, sampling
- laboratory
  - analytical procedures



- all relevant data must be given
- hand out tracer plan as early as possible
- public should be informed
- tracer test plan can be used to ask for permission from authorities
- tracer test plan should be ready at time of proposal to company



- district, local community
- exact description of injection sites
- Home, work, and mobile telephone numbers of participants and substitutes
- relevant authorities
- goal/aim of tracer test
- type and quantity of tracers
- injection time
- quantity of priming and flushing water needed
- list of sampling sites and times

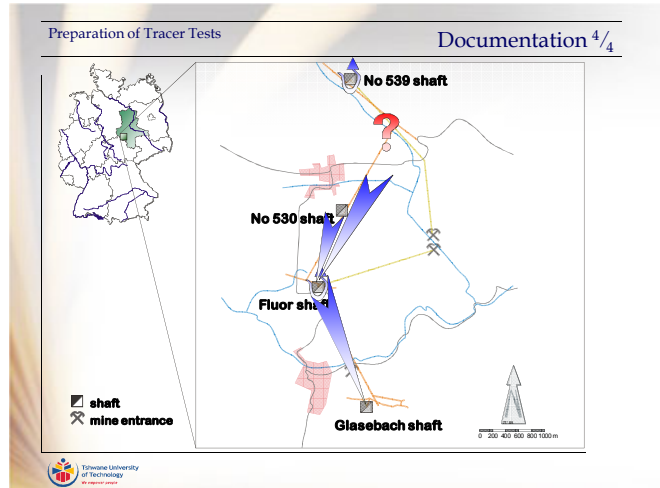
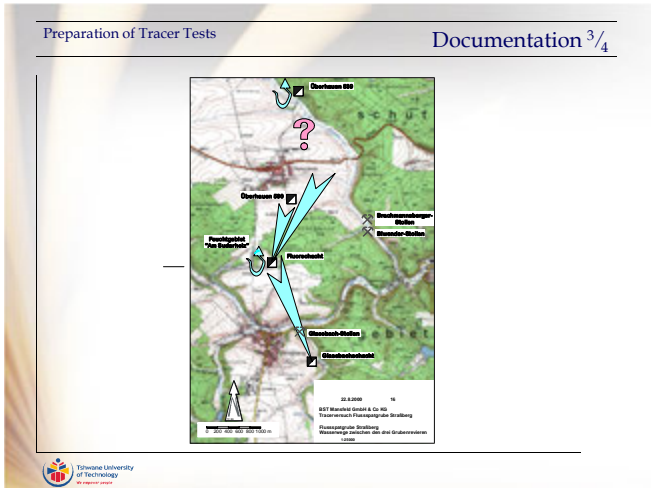


- time of injection
- type of injection
- time of sampling
- sampling procedure
- sampling interval
- sample numbers
  - think about that prior to the start of test
  - Project – Date – Sampling Site – Number
  - BRX-1902-BS1
  - SBG-0708-GS2



- administrative district
- number of topographical map(s)
- detailed description of the investigation area
- name of project head
- type and quantity of tracer
- injection site(s) with co-ordinates
- start and finish times of injection
- priming and flushing water amounts
- data about every sampling site
- data presentation (e.g. breakthrough curves)





- Legislation Legal Questions/Permission
- a tracer test is utilization of water
  - utilization of water needs permission
    - ⇒ artificial tracer tests have to be permitted
    - ⇒ natural tracer tests need no permission
  - permitting authorities
    - water authorities, mining authorities
    - contact authorities before applying for a permit
    - take into account guidelines for use of tracers
      - Germany: UBA-list
  - permission takes 4-6 weeks (sometimes even longer)

- Legislation Legal Questions/Permission
- no special legislation for tracer tests exists
  - nevertheless:
    - national or regional water laws
    - in Germany: „Wasserhaushaltsgesetz“
      - § 1a: General Introduction
      - § 2: Need for Permissions
      - § 3: Utilization
      - § 4: Conditions for Utilization
      - § 7: Permission
  - necessary data for permission
    - kind of tracer, duration, start, place, responsible persons




- Legislation Responsibilities in Canada
- In Canada, inform the following authorities
    - Fisheries and Oceans Canada (DFO)
    - Environment Canada
    - Provincial environment department.
  - Guidelines exist in
    - Alberta
  - Draft Version of Guidelines
    - Quebec
  - Case by Case Decision
    - Newfoundland and Labrador
    - Northwest Territories
    - Nunavut

Legislation Provide the following information

---

- District and place name including map and map references
- Person responsible for conducting tracer test and contact phone numbers
- Customer with full address
- Purpose of the tracer test
- Type of tracer with its exact name, CAS number (Chemical Abstracts Service Registry Number), expected dilution, LD<sub>50</sub> (50% lethal dose) data and MSDS sheets
- Description of injection sites including map
- A list of sampling sites including map and sampling intervals
- Water quantities to flush the injection site and the tracer including where water will be drawn from
- Start of tracer test including start and end of tracer injection
- Work schedule for continuous sampling
- Necessary measurements on site
- Work schedule for analytical laboratory
- Emergency plan including contact phone numbers

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Implementation Implementation

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- Injection Time and Injection Procedure
  - when to inject the tracer
  - how to inject the tracer
- Sampling procedure
  - kind of sampling procedure
- Tracer Preparation, Analyses
  - storage, preparation
  - analytical procedures
- Documentation and Presentation
  - report
  - graphical presentation

---




Implementation Injection: Time and Procedure

---

- injection time
  - results from hydrogeological investigation
  - leave enough time for preliminary preparations on site
  - all personnel and material should have enough time to get to the site
  - start at the beginning of a week to avoid the usual „Friday-late-afternoon“ hurry
  - take into account holidays and working hours in factories or mines
  - inject most effective tracers first (long flow distances)

---




Implementation Injection: Time and Procedure

---

- procedure
  - flush containers several times to inject tracer fully
  - flush tracer into aquifer or unsaturated zone by using excess water
  - tracer has to flow through unsaturated zone first before reaching the aquifer
  - avoid any contamination
  - avoid cross-contamination (e.g. pH-meters)
  - dissolve fluorescent tracer dyes in the laboratory
  - it might be necessary to use boreholes for tracer injection
  - keep in mind: tracer injection changes the groundwater's flow properties

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Implementation Injection: Time and Procedure


---

- procedure
  - avoid slow tracer injection
    - „smearing“ could sham non existing dispersion
  - inject tracer as quick as possible (DIRAC-function)

$$\delta(t) = \begin{cases} 0 & t \neq 0 \\ \infty & t = 0 \end{cases}$$

$$\int_{t_1}^{t_2} dt \delta(t) = 1$$



---



Implementation Sampling procedure


---

- people taking the samples have great responsibility - choose them carefully
- correct labelling is essential
- you must stay to the predefined sampling plan
- inform head of project immediately if any changes occur (e.g. no more tracer visible or vice versa)
- if possible, use automatic sampling devices
- pumps can help



#10 Ingenieurbüro für Geotechnik  
 D-02625 Bötzen - 03591/6771-30
   
 Object: Taucherfriedhof/Bautzen  
 Date: 1998-12-01  
 Time: 17:25  
 Responsibility: Tamme  
 Sample: TFB-0112-01

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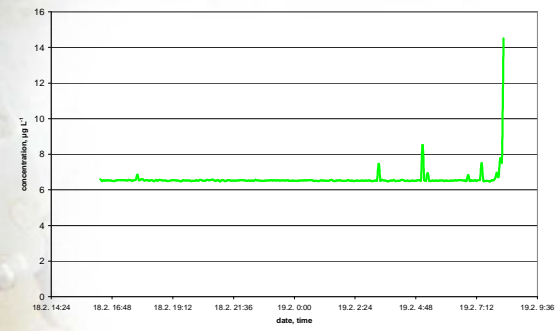
- sample time must be recorded accurately (not as in the sampling plan!)
- sample containers must be clean
- try to use brown glass containers
- store samples in a dark cool place
- take enough sample volume (at least 250 mL)
- samples should be brought to the laboratory as soon as possible



- some minerals have spectral bands similar to the tracers you use
- sort the samples in the order of their extraction to recognize sampling errors
- include background analyses
  - lithium in karst water:  $1 - 2 \mu\text{g L}^{-1}$
  - fluorescein-Na in karst water:  $0.003 \mu\text{g L}^{-1}$
- only if background is known accurately enough, tracer test can be evaluated
- series determination is easier than single determination – but keep in mind tracer decomposition



- background signals



date	time	concentration
18.2.14.24	12:00	6.0
18.2.16.48	13:00	6.0
18.2.19.12	13:00	6.0
18.2.21.36	13:00	6.0
19.2.0.00	13:00	6.0
19.2.2.24	13:00	6.0
19.2.4.48	13:00	6.0
19.2.7.12	13:00	6.0
19.2.9.36	13:00	14.0

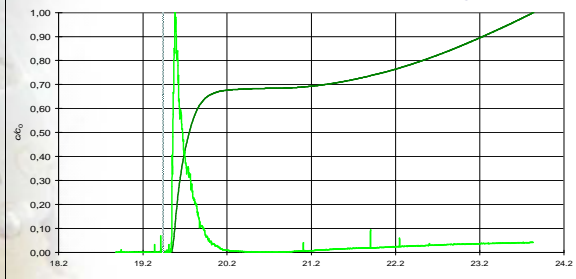
- date
- time
- sampling circumstances
- person in charge of sampling
- weather conditions
- tracer concentration
- physico-chemical parameters

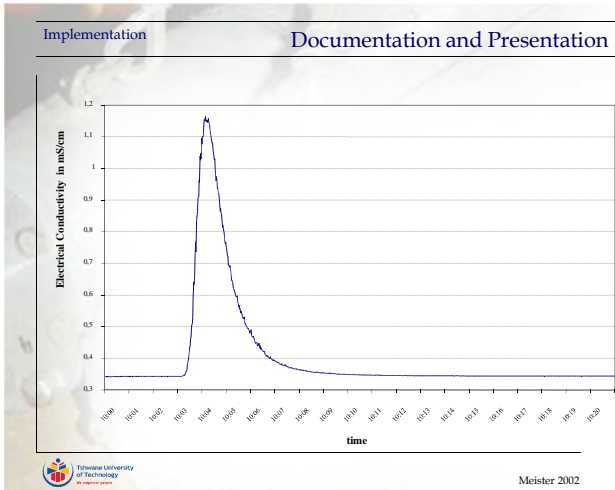
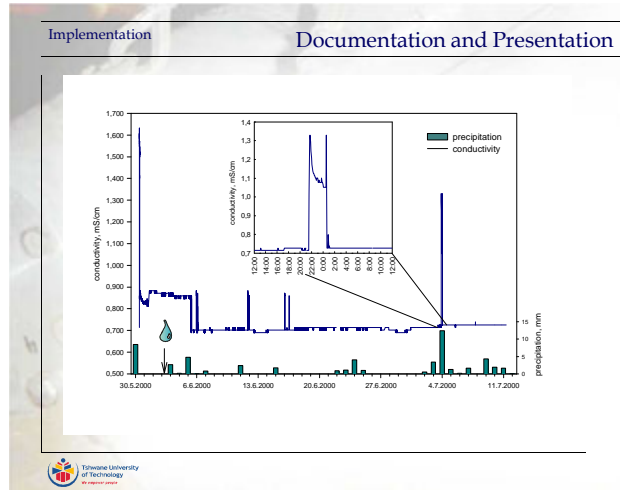
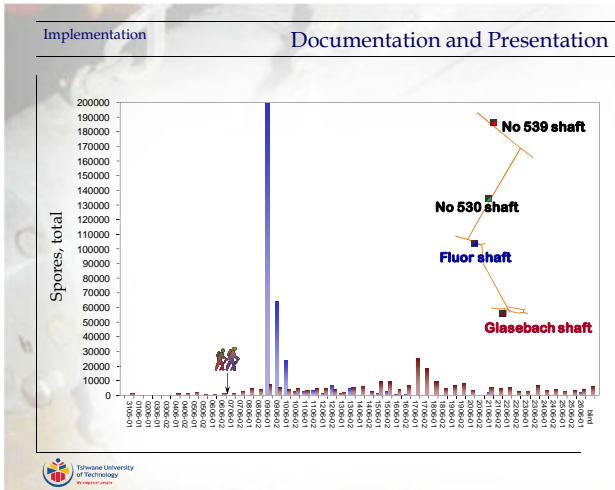


- municipal and administrative district
- number of topographical map
- designation of investigation area
- name of project head
- type and quantity of tracer
- injection sites with name, co-ordinates, altitude, geological formation
- start and finish times of tracer injections, priming and flushing water, flow volume
- sampling sites
  - name, co-ordinates, discharge or pump rate, tracer concentration, start and finish of observation



- use breakthrough curves
- normed curves should be used
- summation curves ease understanding

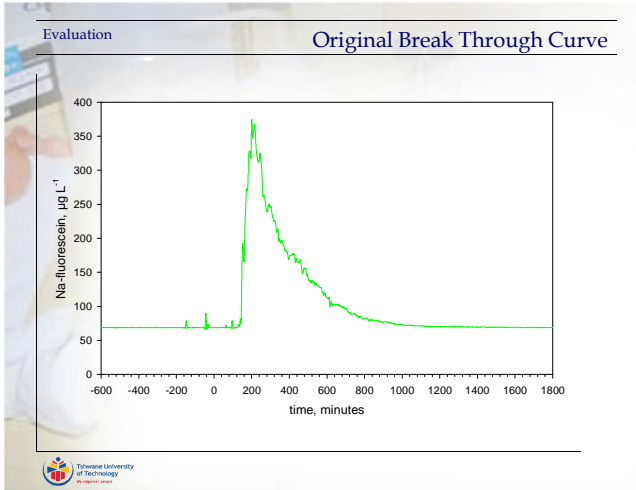
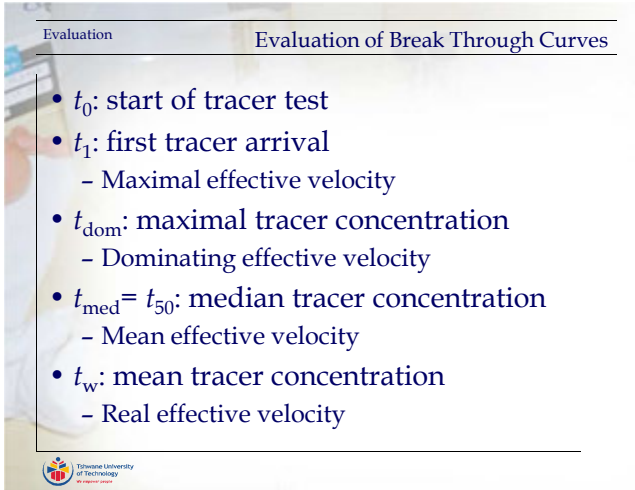
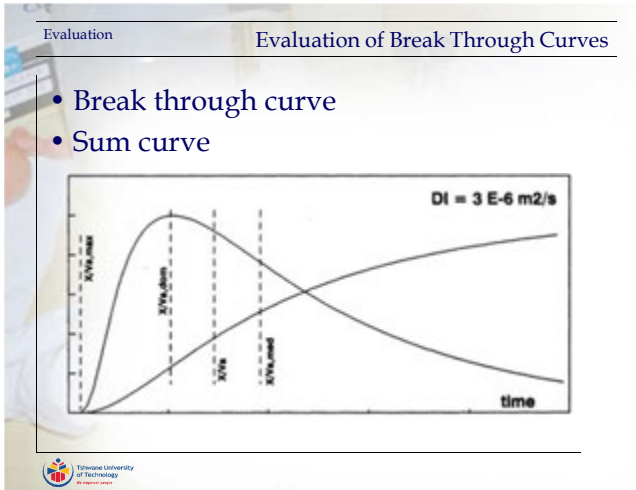
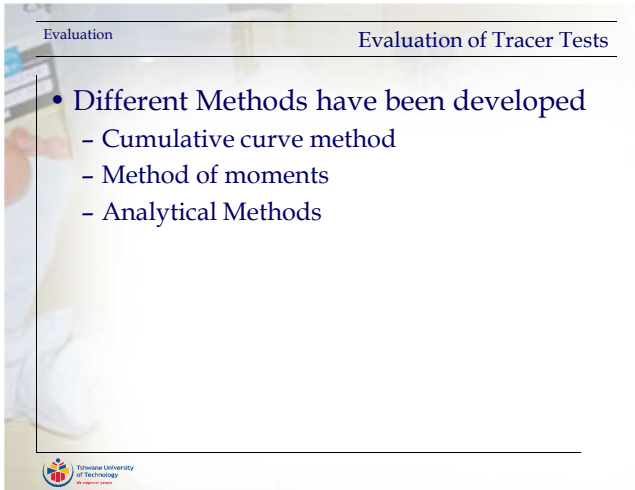
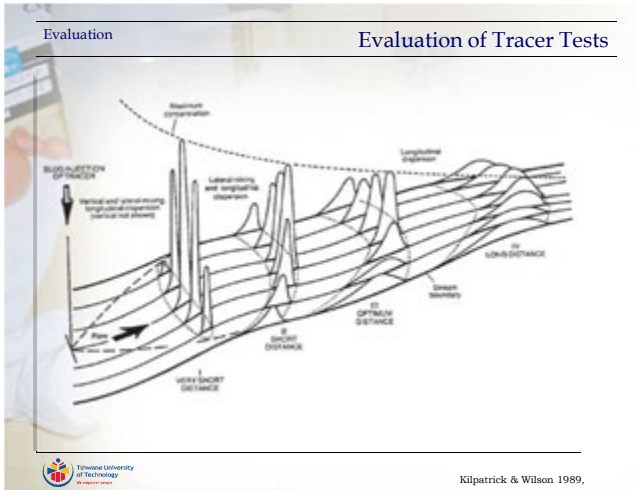
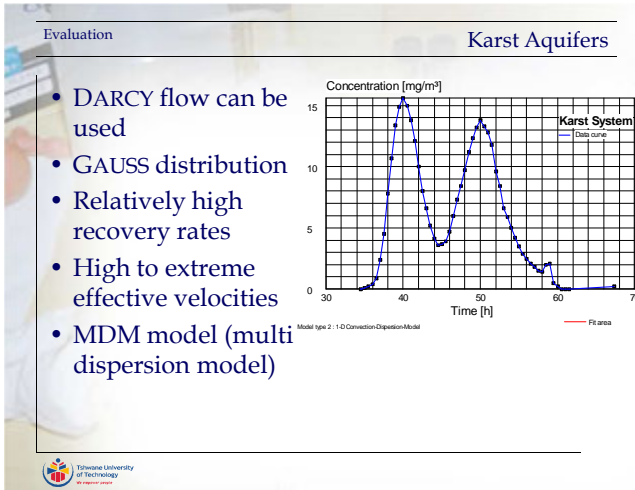


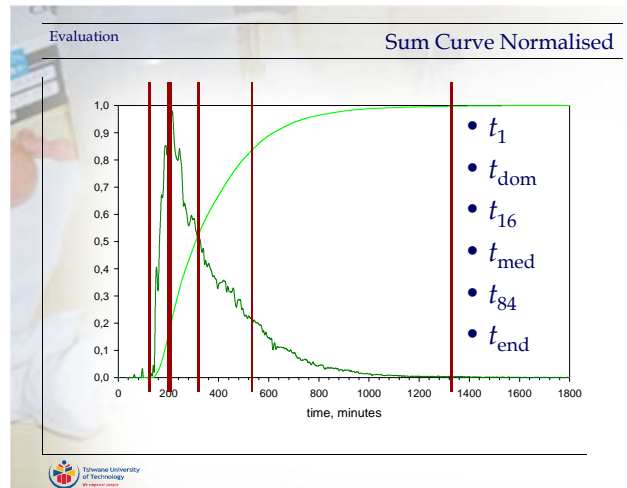
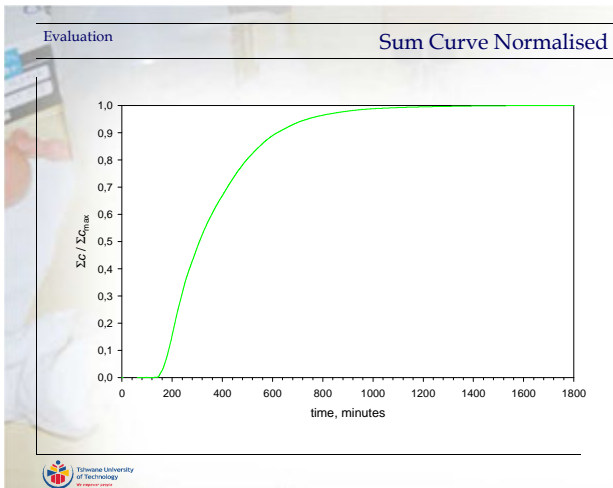
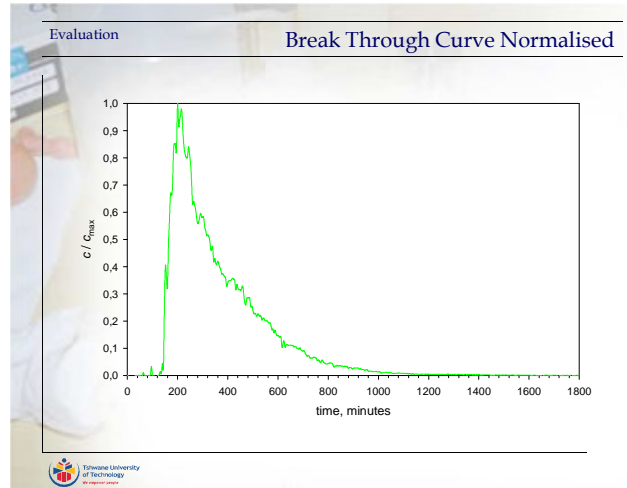
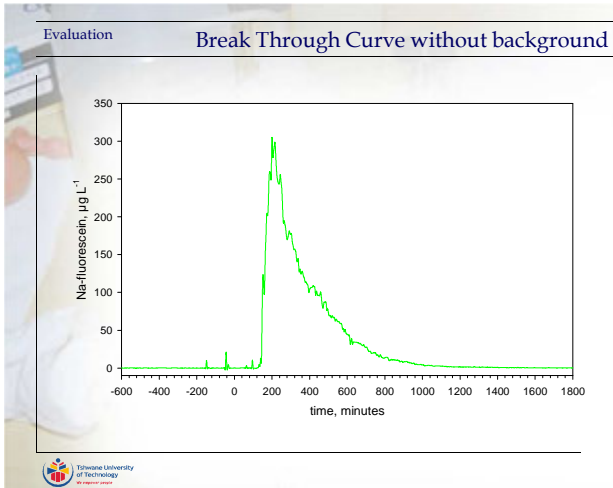


- Evaluation Types of Aquifers and Methods
- Porous Aquifers
  - Fracture Aquifers
  - Karst Aquifers
  - Evaluation of Break Through Curves
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- Evaluation Porous Aquifers
- Darcy flow
  - GAUSS distribution
  - Relatively high recovery rates
  - Low effective velocities
  - Analytical solution is possible
- 
- concentration
- flow path
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- Evaluation Fracture Aquifers
- No Darcy flow
  - No GAUSS distribution
  - Relatively low recovery rates
  - Mean effective velocities
  - SFDM model commonly used (single fissure dispersion model)
- 
- $C(t) B20E$
- T [Days]
- Tishane University of Technology



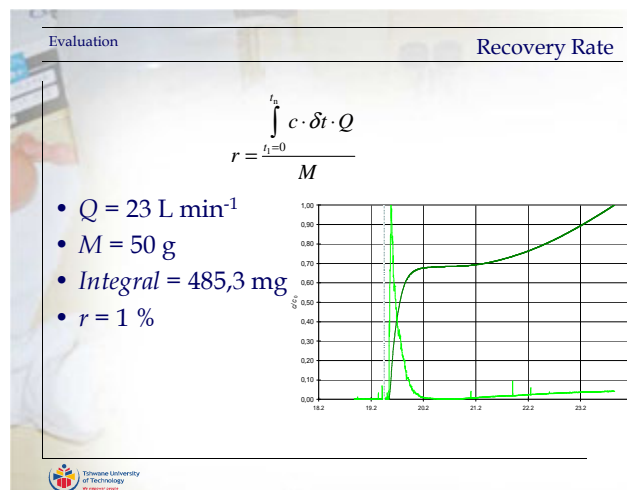


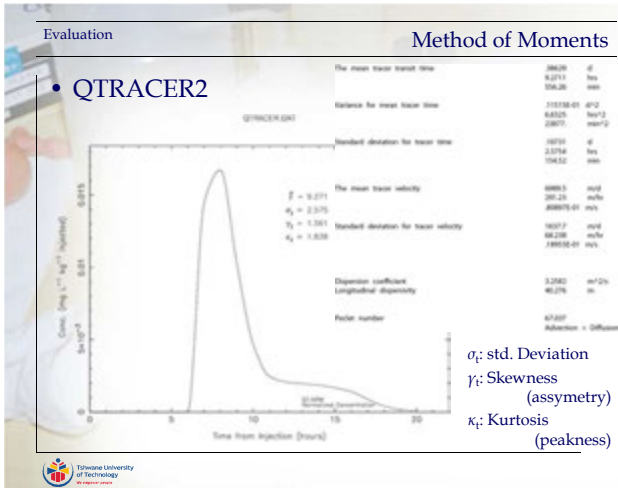
Evaluation Velocities

$$v = \frac{x}{t} \quad D_L = \frac{v_{50}^2 \cdot (t_{84} - t_{16})^2}{8 \cdot t_{50}} \quad D_L = v \cdot \lambda$$

• $t_1$	150 min:	$v = 6,67 \text{ cm min}^{-1}$
• $t_{\text{dom}}$	200 min:	$v = 5,00 \text{ cm min}^{-1}$
• $t_{16}$	201 min:	$v = 4,98 \text{ cm min}^{-1}$
• $t_{\text{med}}$	309 min:	$v = 3,24 \text{ cm min}^{-1}$
• $t_{84}$	538 min:	$v = 1,86 \text{ cm min}^{-1}$
• $t_{\text{end}}$	1500 min:	$v = 0,67 \text{ cm min}^{-1}$
• $D_L = 1,5 \text{ m}^2 \text{ min}^{-1}$		$(\lambda = 0,005 \text{ m})$
• $D_T \approx 0,1 D_L = 0,15 \text{ m}^2 \text{ min}^{-1}$		

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- Examples Examples
- Surface/Ground Water
    - Surface flow measurements
    - hydraulic connection (Taucherfriedhof Bautzen)
  - Mine Water
    - Multiple shafts (Straßberg/Harz)
    - Single shaft (Brixlegg/Tirol)

- Examples Surface Flow Measurements
- Standing (lakes) and flowing (rivers) water bodies
  - Detection of flow connections
  - Flow Velocities
  - Mixing conditions
  - Dispersion
  - Flow Measurements
  - Evaluation of physical parameters
    - Oxygen depletion (degasing measurements)
    - Light input (decomposing tracer)

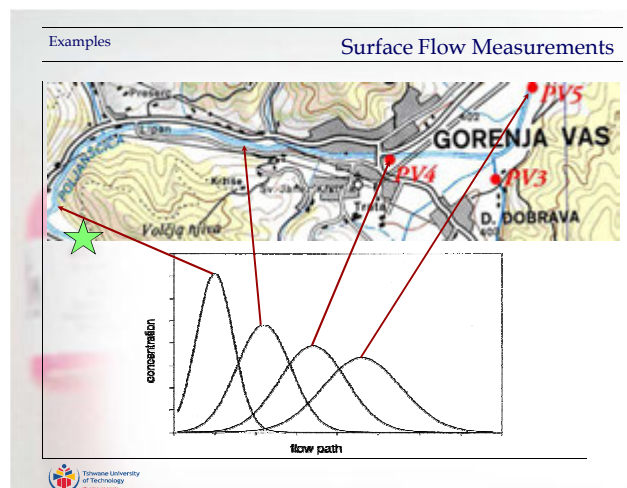
Examples Surface Flow Measurements

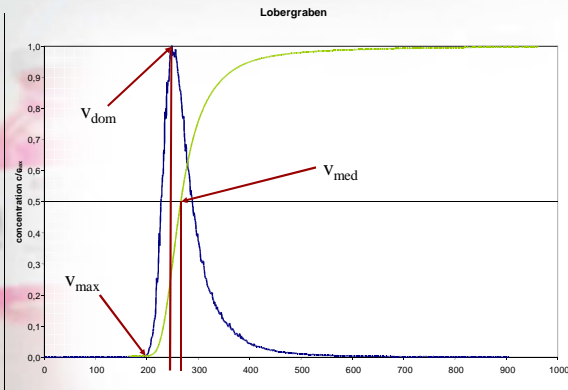
- Flow velocities
- Known tracer amount injected
- Mixing of tracer
- DIRAC impulse

$$Q = \frac{M}{\int_{t=0}^{t=\infty} c_t dt}$$

$M$  = injected tracer amount  
 $C_t$  = concentration of tracer at time  $t$   
 $t$  = time of tracer sampling

- Examples Surface Flow Measurements
- Measurement in middle of river
  - Possible Tracers
    - NaCl
    - Na-fluorescein (Uranine)
    - Rhodamine WT
  - Fluorimeter (fibre optics)
  - Electrical Conductivity





• Results

$$Q = \frac{M}{\int_{t=0}^{t=\infty} c_t dt} = \frac{10,000 \text{ g}}{65.9 \frac{\text{g}}{\text{L}}} = 0.15 \text{ L} \cdot \text{s}^{-1} = 9.1 \text{ L} \cdot \text{min}^{-1}$$

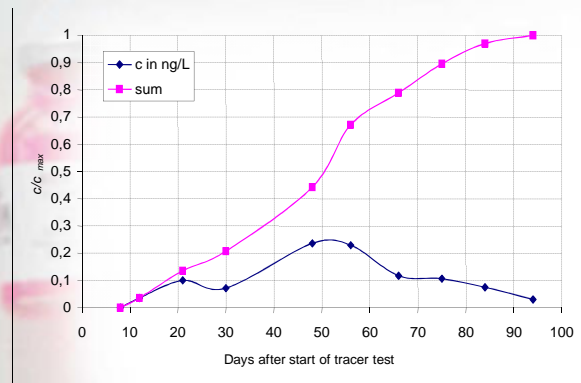
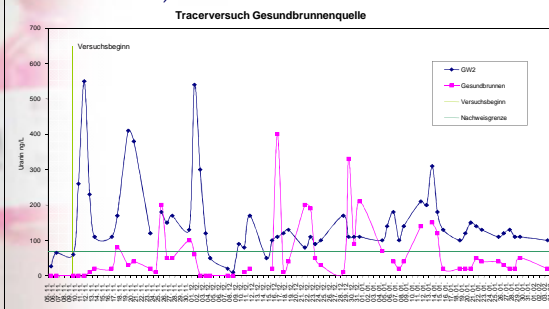
$M$  = injected tracer amount

$c_t$  = concentration of tracer at time  $t$

$t$  = time of tracer sampling

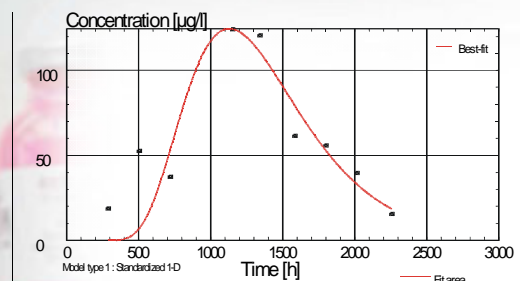


• Taucherfriedhof Bautzen (hydraulic connection)



- $t_1 = 12 \text{ d} \Rightarrow v_1 = 2.7 \cdot 10^{-4} \text{ m} \cdot \text{s}^{-1}$
- $t_{\text{dom}} = 49.5 \text{ d} \Rightarrow v_{\text{dom}} = 6.5 \cdot 10^{-5} \text{ m} \cdot \text{s}^{-1}$
- $t_{16} = 25.6 \text{ d} \Rightarrow v_{16} = 1.3 \cdot 10^{-4} \text{ m} \cdot \text{s}^{-1}$
- $t_{50} = 49.7 \text{ d} \Rightarrow v_{50} = 6.5 \cdot 10^{-5} \text{ m} \cdot \text{s}^{-1}$
- $t_{84} = 68.5 \text{ d} \Rightarrow v_{84} = 4.7 \cdot 10^{-5} \text{ m} \cdot \text{s}^{-1}$

$$v_a = \frac{v_f}{n_e} = \frac{2.1 \cdot 10^{-6}}{0.025} = 8.4 \cdot 10^{-5} \text{ m} \cdot \text{s}^{-1}$$



$$t_{50} = 56.9 \text{ d} \Rightarrow v_{50} = 5.7 \cdot 10^{-5} \text{ m/s}$$

$$D_L = 5 \cdot 10^{-3} \text{ m}^2/\text{s}$$

$$\lambda = 91 \text{ m}$$



- Brixlegg/Tirol
  - historical silver-barite mine
  - 100 m deep shaft
    - 20, 40, 70, 100 m levels
  - Flooded since 1990
  - Outflow  $20-50 \text{ L} \cdot \text{min}^{-1}$

